



## Preparation of *Alpinia galanga* Water Extract with High Antioxidant Properties

Nik Hasan, M.K\*., Kamarazaman, I.S., Azman, M., Abd Rashid L.

*Natural Product Division, Forest Research Institute Malaysia (FRIM), 52109 Kepong, Selangor Darul Ehsan*

\*For correspondence: mohdkamal@frim.gov.my

---

**Abstract:** *Alpinia galanga* is one of the most common herbs in Malaysia. Because of its good aroma and taste, *A. galanga* has been used widely for Asian cooking. The objective of this study is to determine the optimum parameter (temperature and period) for *A. galanga* water extract preparation with preservation of its antioxidant property. *A. galanga* was cut, dried and ground into powder. The powder was put into a tube containing water and heated at different temperatures which were 40, 60, 80, and 100°C. Each of temperature, the extract was used with different periods which were 15 min, 30 min, 1 h, 2 h and 3 h. the extract was filtered and used for DPPH, FRAP, TPC and MDA tests to determine its antioxidant value. The result show that *A. galanga* has different antioxidant property when it is prepared with different parameters. It can be concluded that the optimum parameter to prepare *A. galanga* extract with high antioxidant activity is at a the temperature of 80°C with a period of 2 h.

© 2020, Asian Society of Pharmacognosy. All Rights Reserved.

*Keyword: Alpinia galangal; antioxidant*

---

### INTRODUCTION

*Alpinia galanga* (L.) Willd. (Zingiberaceae) known as Galanga, is a member of the ginger family, which originated from Southern China and Thailand (Daniel, 2006, Pillai *et al.*, 2018). The rhizome of Galanga traditionally used in Malaysia to treat fever, headache, cough, respiratory illness, stomachache, indigestion, flatulence, laxation, nausea, vomiting, diarrhea, menstrual pain, skin infection, ringworm, afterbirth and postpartum infection (Burkill, 1935, Basri *et al.*, 2017). Galanga has also exhibited numerous pharmacological activities such as anti-microbial, anti-inflammatory, aphrodisiac, eczema, bronchitis, chest pain, dyspepsia, fever, disease, tumors, diabetes and even HIV (Elyani & Risandiansyah, 2017; Verma *et al.*, 2014; Shetty & Monisha, 2015; Ramesh, 2011). Phytochemical screening of Galanga showed the presence of terpenoids, essential oils, phenolic compounds (phenylpropanoids), and flavonoids (Alajmi *et al.*, 2018). *A. galanga* has been reported to contain alpinin, galangin, methyl cinnamate, 1'-acetoxychavicol acetate, 1'-hydroxychavicol acetate,  $\alpha$ -humulene, limonene, myrcene,  $\alpha$ -pinene,  $\beta$ -pinene, quercetin, kaempferol, quercetin, and 3-methyl ether (Janssen *et al.*, 1985; Nori *et al.*, 1988; De Pooter *et al.*, 1985). A compound isolated from Galanga, 1'S-1'-acetoxychavicol acetate (ACE), was so far the major compound reported with various biological activities (Baradwaj, 2017). An antioxidant is substances or compounds, when presence even in a low concentration, can inhibit



or delay the oxidation of other substances. Oxidation is the chemical reaction that produces free radicals, leading to the chain reactions that may damage cells, lipids, proteins or DNA (Lyras et al., 1997). When there is an unbalance between free radicals and antioxidants, in favor of the former, a condition called oxidative stress occur. Oxidative stress may result in various health conditions including heart diseases, cancer, stroke, arthritis, Parkinson's disease, and other inflammatory or ischemic conditions (Jenner, 2003; Sayre et al., 2001; Sosa et al., 2013). Several assays have been used to determine the antioxidant activity of plant extracts including DPPH scavenging assay, total phenolics content (TPC) determination, ferric reducing antioxidant power (FRAP), and malondialdehyde (MDA) assays. However, the antioxidant properties of plant extracts can be inconsistent among different assays tested. The inconsistency can be due to the different mechanisms involved in the assay itself. For instance, FRAP assay determines the reducing capability based on ferric ion and DPPH assay determines the scavenging effect of antioxidants towards stable radicals, DPPH. There is no single method that is comprehensive enough to determine the antioxidant properties of plant extracts. Therefore, the use of more than one method is recommended (Prior et al., 2005). Different extraction temperatures and periods may affect the antioxidant properties of plant extract. There were several reports which indicate the effect of different extraction temperatures toward antioxidant properties of plant extracts and natural products (Reblova, 2012; Hossain et al., 2013; Molaveisi et al., 2019). This study was performed to determine the best extraction temperature and extraction period to produce standardized *Alpinia galanga* extract.

## PROTOCOL AND METHODS

*Crude extraction:* First, the root of *A. galanga* was washed and cut into small pieces. Then, *A. galanga* was put into the oven for the drying process at a temperature of 55°C for two days. After that, *A. galanga* was ground into crude powdered form.

*Water extraction:* 1.77 g of *A. galanga* was weighed by using an electronic balance and put into glass tubes. After that, the glass tube was added with 10 ml of distilled water. Then, it was heated different temperatures (40, 60, 80, 100 °C) for different times (15 min, 30 min, 1 h, 2 h, and 3 h). The extract was filtered and put into a labeled tube.

*Diphenyl picrylhydrazil (DPPH) assay:* The effect of extract of extracts on DPPH radical was estimated using the method of Yen and Hsieh (1998). The positive controls were BHT (Butylated Hydroxytoluene) and Vitamin C (Ascorbic Acid). The wavelength that was used was 540nm.

*Ferric Reducing Antioxidant Power (FRAP) assay:* The ferric reducing ability of the plant materials was assessed using the method described by Benzie and Strain (1996). The absorbance was measured spectrophotometrically at 595nm. The final results were expressed as mol Fe (II) equivalent/g sample in the fresh sample.

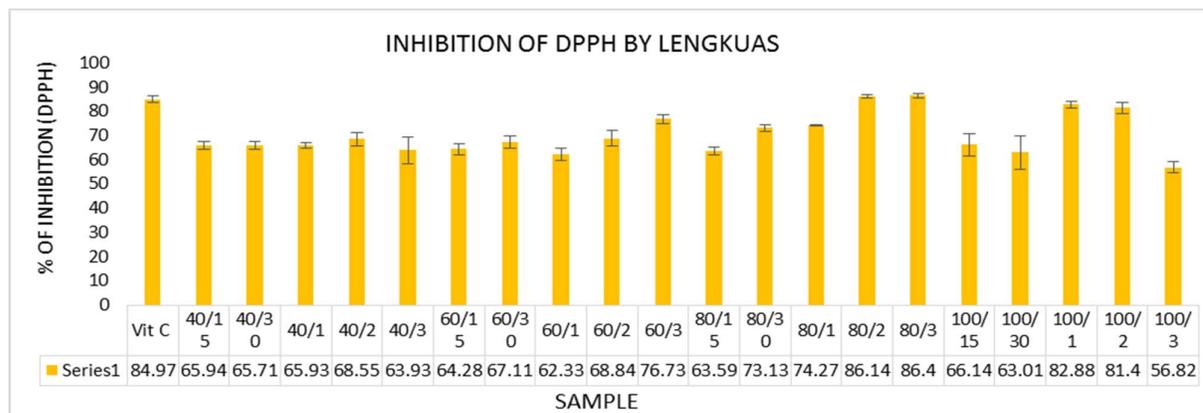
*TPC assay (Total Phenolic Content):* The concentration of total phenolic was based on the method described by Velioglu et. al. (1998). The absorbance of the reaction was measured at 595nm against a blank.

*MDA assay (Malondialdehyde):* A modified thiobarbituric acid-reactive species (TBARS) assay was used to measure the lipid peroxide formed, using egg-yolk homogenates as lipid-rich media (Ohkawa et al., 1979). 250 µL of Egg homogenate and 50 µL of extract were mixed in a test tube and the volume was made up to 500 µL, by adding distilled water. Finally, 25 µL “FeSO<sub>4</sub>” (0.07 M) was added to the mixture and incubated for 30 min. Thereafter, 750 µL of 20% acetic acid (pH 3.5) and 750 µL of 0.8% TBA (w/v) (prepared in 1.1% sodium dodecyl sulfate) and 25

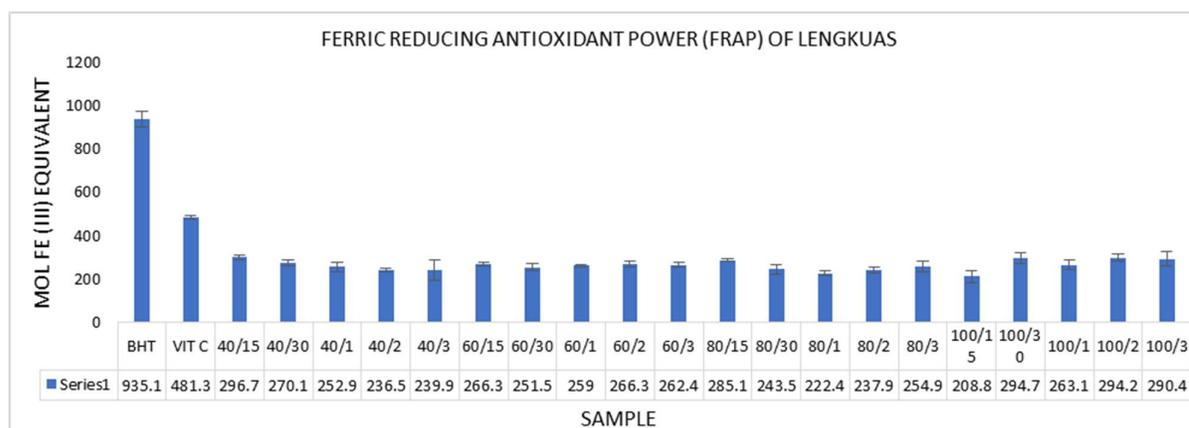


$\mu$ L 20% TCA was added, vortexed, and then heated in a boiling water bath for 60 min. After cooling, 3.0 mL of 1-butanol was added to each tube and centrifuged at 3000 rpm for 10 min. The absorbance of the organic upper layer was measured against 3 mL butanol at 532 nm.

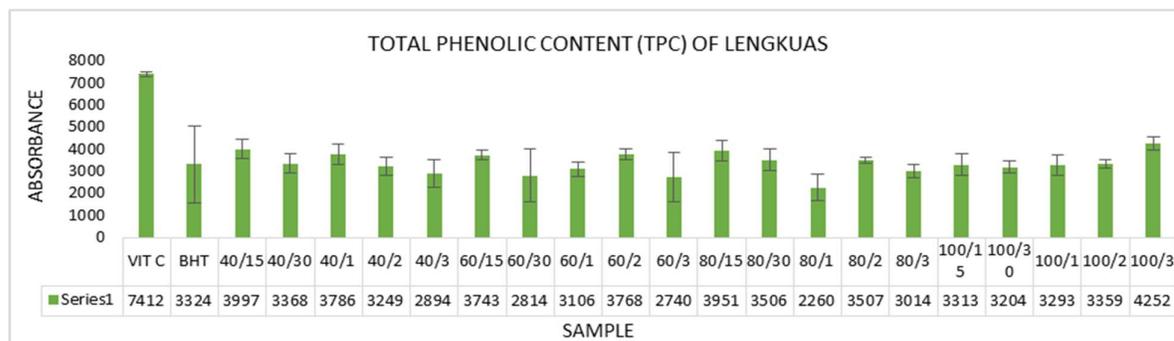
## RESULTS



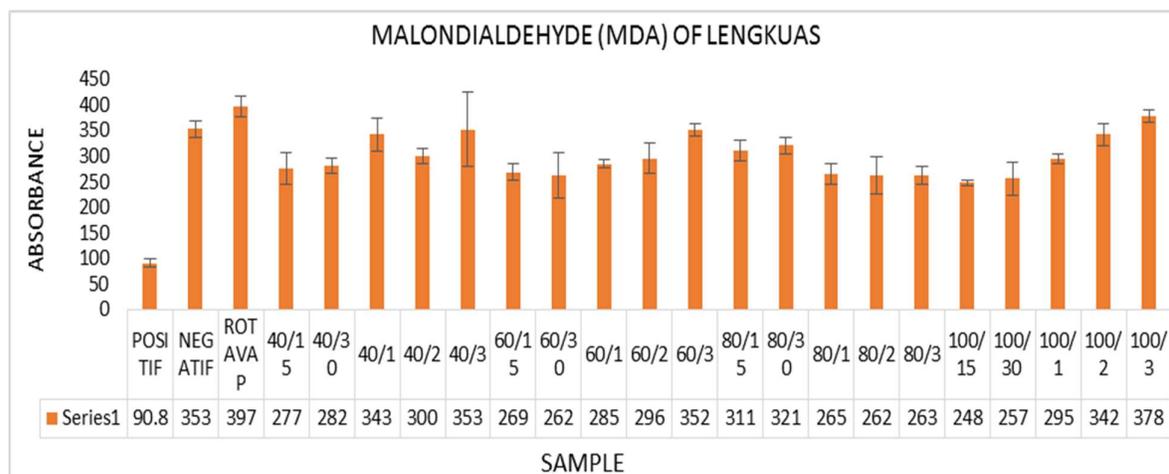
**Graph 1:** Based on graph 1, it can be seen that the temperature of 80°C with 2 hours period shows a consistent DPPH inhibition percentage compare to others.



**Graph 2:** Based on graph 2, it can be seen that all parameter showed lower FRAP value compared to positive control which was BHT and Vitamin C.



**Graph 3:** Based on graph 3, it can be seen that all parameter showed lower TPC value compared to positive control BHT. However, there were several parameters were not significant compared with Vitamin C.



**Graph 4:** Malondialdehyde (MDA), a secondary product of the oxidation of polyunsaturated fatty acids, reacts with two molecules of thiobarbituric acid (TBA), yielding a pinkish-red chromogen with an absorbance maximum at 532 nm (Janero, 1990). Reducing of MDA level showed the sample can decrease the oxidative activity. The lowest MDA level was 100°C/15h. The extract that was prepared using parameter 80°C/2h also showed lower MDA level compared to the negative control.

## CONCLUSION

In the DPPH and MDA test, the most effective parameter that showed a higher value of antioxidant activity was 80°C at 2 hours. Besides that, the extract can reduce MDA concentration compared to the negative control.



## ACKNOWLEDGMENTS

This work was financially supported by the RMK11 Project, Natural product Division, Cell Signalling and In Vivo Research Laboratory.

## DECLARATION OF CONFLICT OF INTEREST

The authors declare that there are no conflicts of interest.

## REFERENCES

- Alajmi, M.F., Mothana, R.A., Al-Rehaily, A.J. & Khaled, JM. Antimycobacterial Activity and Safety Profile Assessment of *Alpinia galanga* and *Tinospora cordifolia* Evidence-Based Complementary and Alternative Medicine Volume 2018,
- Baradwaj RG, Rao MV, Senthil Kumar T. Novel purification of 1'S-1'-Acetoxychavicol acetate from *Alpinia galanga* and its cytotoxic plus antiproliferative activity in colorectal adenocarcinoma cell line SW480. *Biomed Pharmacother.* 2017;91:485-93. DOI:10.1016/j.biopha.2017.04.114
- Basri A.M., Taha H., and Ahmad N., A Review on the Pharmacological Activities and Phytochemicals of *Alpinia officinarum* (Galanga) Extracts Derived from Bioassay-Guided Fractionation and Isolation, *Pharmacogn Rev.* 2017 Jan-Jun; 11(21): 43–56.
- Benzie I.F.F., Strain, J.J. (1996). Ferric reducing the ability of plasma (FRAP) as a measure of antioxidant power: The FRAP assay. *Anal Biochem*, 239:70-76.
- Burkill IH. A dictionary of the economic products of the Malay Peninsula. Vol. 2. London: Published on behalf of the Governments of the Straits Settlements and the Federated Malay States by the Crown Agents for the Colonies. 1935; p.1306-1310.
- Daniel M. Medicinal Plants: Chemistry and Properties. Enfield: Science Publishers; 2006. p. 63.
- De Pooter HL, Omar MN, Ciiksaet BA, Schamp NM. The essential oil of greater galanga (*Alpinia galanga*). *Phytochemistry.* 1985;24:93-96.
- Elyani, H. & Risandiansyah, R. "Antibacterial potential of four herbal plants (*Syzygium cumini*, *Piper ornatum*, *Anredera cordifolia*, and *Alpinia galanga*) against *Staphylococcus aureus* and *Escherichia coli*," *JIMR-Journal of Islamic Medicine Research*, vol. 1, no. 2, 2017.
- Janero DR. Malondialdehyde and thiobarbituric acid-reactivity as diagnostic indices of lipid peroxidation and peroxidative tissue injury. *Free Radical Biology and Medicine.* 1990;9(6):515–540.
- Janssen AM, Scheffer JJC. Acetoxychavicol acetate, an antifungal component of *Alpinia galanga*. *Planta Med.* 1985;51 (6):507-511.
- Jenner P. Oxidative stress in Parkinson's disease. *Ann Neurol.* 2003;53: S26–S36. 8.
- Lyras L, Cairns NJ, Jenner A, Jenner P, Halliwell B. An assessment of oxidative damage to proteins, lipids, and DNA in the brain from patients with Alzheimer's disease. *J Neurochem.* 1997;68:2061–2069.
- Mohammad A Hossain\*, Zawam Hamood AL-Mijizy, Kawther Khalifa Al-Rashdi, Afaf M Weli, Qasim Al-Riyami Effect of temperature and extraction process on antioxidant activity of various leaves crude extracts of *Thymus vulgaris* *Journal of Coastal Life Medicine* 2013; 1(2): 130-134
- Molaveisi, M. Beigbabaei, A., Akbari, E., Noghabi, M.S. & Mohamadi, M. Kinetics of temperature effect on antioxidant activity, phenolic compounds and color of Iranian jujube honey *Heliyon.* 2019 5(1): e01129.
- Nori T, Sekiya T, Katoh M, et al. Inhibitors of xanthine oxidase from *Alpinia galanga*. *Chem Pharm Bull.* 1988;36:244-248.
- Ohkawa H, Ohishi N, Yagi K, Assay for lipid peroxides in animal tissues by the thiobarbituric acid reaction, *Anal Biochem.* 1979 Jun; 95(2):351-8.
- Pillai MK, Young DJ, Bin Hj Abdul Majid. Therapeutic Potential of *Alpinia officinarum*. *Mini Rev Med Chem.* 2018;18(14):1220-1232.
- Prior, R.L.; Wu, X.; Schaich, K. Standardized Methods for the Determination of Antioxidant Capacity and Phenolics in Foods and Dietary Supplements. *J. Agric. Food Chem.* 2005, 53 (10),4290–4302.
- Ramesh KV, Garima S, Pradeep S, Jha KK, Khose RL. *Alpinia galanga* an important medicinal plant: a review, *Der Pharm. Sin.* 2011;2(1):142-54
- Réblová, Z. Effect of Temperature on the Antioxidant Activity of Phenolic Acids *Czech J. Food Sci.* Vol. 30, 2012, No. 2: 171–177
- Sayre LM, Smith MA, Perry G. Chemistry and biochemistry of oxidative stress in neurodegenerative disease. *Curr Med Chem.* 2001;8:721–738. 10. Toshniwal PK, Zarling EJ. Evidence for increased lipid peroxidation in multiple sclerosis. *Neurochem Res.* 1992;17:205–207.
- Shetty G. R. & Monisha, S. "Pharmacology of an endangered medicinal plant *Alpinia galanga*-a review," *Research Journal of Pharmaceutical, Biological and Chemical Sciences*, vol. 6, no. 1, pp. 499–511, 2015.



**ASIAN JOURNAL OF PHARMACOGNOSY**

*Asian J. Pharmacogn* 4(1): 43-48

© 2020, Asian Society of Pharmacognosy. All Rights Reserved.

ISSN-0128-1119

Sosa V, Moliné T, Somoza R, Paciucci R, Kondoh H, LLeonart ME. Oxidative stress and cancer: an overview. *Ageing Res Rev.* 2013 Jan;12(1):376-90.

Velioglu, Y.S., Mazza, G., Gao, L., Oomah, B.D. (1998). Antioxidant activity and total phenolics in selected fruits, vegetables, and grain products. *J. Agric. Food Chem*, 46:4113–4117.

Verma, R. Mishra, G. Singh, P. Jha, K. & Khosa, R. “*Alpinia galanga*—An important medicinal plant: a review,” *Der Pharmacia Sinica*, vol. 2, no. 1, pp. 142–154, 2011.

Yen, G.C, Hsieh, C.L. (1998). Antioxidant activity of extracts from *Du-Zhong* (*Eucoma ulmoides*) toward various lipid peroxidation models in vitro. *Journal of Agricultural and Food Chemistry*, 46: 3952-3957.